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Neuroanatomical Changes in People with High Schizotypy: Relationship to Glutamate Levels

Abbreviated title: Glutamate and Structure in Schizotypy

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ABSTRACT

Background: Cortical glutamatergic dysfunction is thought to be fundamental for psychosis development, and may lead to structural degeneration through excitotoxicity. Glutamate levels have been related to gray matter volume (GMV) alterations in people at ultra-high risk of psychosis, and we previously reported GMV changes in individuals with high schizotypy (HS), which refers to the expression of schizophrenia-like characteristics in healthy people. This study sought to examine whether GMV changes in HS subjects are related to glutamate levels.

Methods: We selected twenty-two healthy subjects with HS and 23 healthy subjects with low schizotypy (LS) based on their rating on a self-report questionnaire for psychotic-like experiences. Glutamate levels were measured in the bilateral anterior cingulate cortex (ACC) using proton magnetic resonance spectroscopy, and GMV was assessed using voxel-based morphometry.

Results: Subjects with HS showed GMV decreases in the rolandic operculum/superior temporal gyrus ($p_{FWE} = 0.045$). Significant increases in GMV were also detected in HS, in the precuneus ($p_{FWE} = 0.043$), thereby replicating our previous finding in a separate cohort, as well as in the ACC ($p_{FWE} = 0.041$). While the HS and LS groups did not differ in ACC glutamate levels, in HS subjects ACC glutamate was negatively correlated with ACC GMV ($p_{FWE} = 0.026$). Such association was absent in LS.

Conclusions: Our study shows that GMV findings in schizotypy are related to glutamate levels, supporting the hypothesis that glutamatergic function may lead to structural changes associated with the expression of psychotic-like experiences.

INTRODUCTION

Reductions in gray matter volume have been extensively reported in patients with schizophrenia, predominantly in temporal, frontal and parietal brain regions (Bora *et al.*, 2011, Chan *et al.*, 2011, Haijma *et al.*, 2013). Such volumetric decreases are also found, albeit attenuated, in unaffected relatives of schizophrenia probands (Boos *et al.*, 2007, McDonald *et al.*, 2008), and in subjects at ultra-high risk (UHR) for psychosis (Fusar-Poli *et al.*, 2011). In UHR individuals, gray matter volume reductions in medial temporal regions are shown to predict subsequent transition to psychosis (Mechelli *et al.*, 2011) and, longitudinally, an accelerated thinning of frontal areas has been associated with progression to psychosis (Cannon *et al.*, 2015). However, the neurobiological mechanisms underlying volumetric changes in patients with psychosis and in people with subclinical psychotic-like experiences are unclear.

A possible mechanism for neuroanatomical changes in psychosis is hypofunction of the N-methyl D-aspartate (NMDA) glutamate receptor (Abbott and Bustillo, 2006, Deutsch *et al.*, 2001). NMDA-receptor dysfunction is found to lead to insufficient excitation of GABAergic inhibitory interneurons, downregulating their activity and consequently increasing activity of pyramidal glutamatergic neurons; this may in turn result in excessive glutamate concentrations around neurons and potentially in excitotoxicity through the influx of calcium ions (Choi, 1994). Thus, excessive glutamate is proposed as a mechanism that would give rise to alterations in gray matter volume in schizophrenia and groups at risk of developing the disorder (Port and Agarwal, 2011).

Increased concentrations of glutamatergic metabolites are observed in schizophrenia across several cortical areas (Marsman *et al.*, 2013, Merritt *et al.*, 2016), and prefrontal elevations in levels of Glx (glutamate + glutamine) are found in antipsychotic-

naïve individuals at UHR for psychosis (Merritt *et al.*, 2016), highlighting a role for abnormal levels of glutamatergic metabolites in increasing psychosis vulnerability. Currently, however, human imaging studies provide only limited evidence to support the hypothesis that increased glutamate signaling may lead to excitotoxicity and reduced gray matter volume. In support of this hypothesis, there are negative relationships between caudate Glx levels and caudate volume in first episode psychosis (Plitman *et al.*, 2016), left hippocampal Glx levels and left hippocampal volume in unmediated patients (Kraguljac *et al.*, 2013), and thalamic glutamate levels and cingulate cortex volume in UHR subjects (Stone *et al.*, 2009). In contrast, positive relationships between thalamic glutamate metabolite levels and more widespread volumetric reductions have also been reported (Aoyama *et al.*, 2011, Stone *et al.*, 2009, Theberge *et al.*, 2007). Given that brain glutamate levels can be affected by antipsychotic treatment (Egerton *et al.*, 2017), further evidence in antipsychotic-naïve individuals may help clarify the relationship between glutamate levels and gray matter changes associated with psychosis.

Consistent with a continuum model of psychosis, which proposes dimensional continuity between subclinical psychotic-like experiences in healthy individuals (or *schizotypy*) and in clinically relevant psychosis (Linscott and van Os, 2013, Nelson *et al.*, 2013), volumetric changes have also been documented in schizotypy (Modenato and Draganski, 2015). While the majority of healthy people with schizotypy as identified through self-report questionnaires are not expected to develop psychosis, high schizotypy is associated with higher risk for developing a psychotic disorder (Kwapil *et al.*, 2013), and represents a useful and widely applied paradigm to investigate neurobiological factors associated with schizophrenia spectrum disorders (Barrantes-Vidal *et al.*, 2015). High schizotypy involves typically non-treatment-seeking individuals from the general population who are commonly identified through psychometrically validated self-report measures, such

as the Oxford and Liverpool Inventory of Feelings and Experiences (O-LIFE) questionnaire (Mason *et al.*, 2005), or the Schizotypal Personality Questionnaire (SPQ) (Raine, 1991). After our initial report of increased gray matter volume in posterior cortical midline areas in high schizotypy (Modinos *et al.*, 2010), subsequent studies found decreases in other fronto-temporal regions including the anterior cingulate cortex (Ettinger *et al.*, 2012, Nenadic *et al.*, 2015, Satterthwaite *et al.*, 2016, Wang *et al.*, 2015), while also a positive association between posterior cortical midline volume and schizotypy scores was reported (Nenadic *et al.*, 2015). Furthermore, a more recent study described both negative and positive associations between gray matter volume and the degree of schizotypy, with negative correlations involving frontal and temporal lobes and positive correlations comprising parietal, temporal and subcortical regions (Wiebels *et al.*, 2016). With regard to glutamate levels in schizotypy, we recently reported no differences between high and low schizotypes in the anterior cingulate cortex, although the striking finding in this group was that glutamate levels were associated with increased neural response to emotion in the striatum, medial prefrontal cortex and amygdala (Modinos *et al.*, 2017). Since replication studies are infrequent in neuroimaging research, and considering that the high schizotypy paradigm allows investigation of neurobiological processes on the psychosis continuum without confounding effects of exposure to previous antipsychotic treatment or illness chronicity, our study sought to address the issue of whether the predicted alterations in gray matter volume in high schizotypy subjects would be related to glutamate levels in the anterior cingulate cortex. The cingulate cortex was chosen given that abnormalities in glutamatergic metabolites in this region are particularly apparent in the at-risk stage of psychosis (Merritt *et al.*, 2016), and that altered anterior cingulate volume has been found in schizotypy (Ettinger *et al.*, 2012). We tested the hypothesis that volumetric changes in high schizotypy individuals would be negatively correlated with glutamate levels in the anterior cingulate cortex.

MATERIALS AND METHODS

Participants

The recruitment procedure is described in detail in our recent article (Modinos *et al.*, 2017). Briefly, two-hundred and fifty healthy volunteers who responded to the Research Volunteer Recruitment Webpage of King's College London were pre-screened with the short version of the O-LIFE questionnaire (Mason *et al.*, 2005). Participants with high scores on the Unusual Experiences (UE) subscale of the O-LIFE (UE score >7, *HS* group), and participants with low levels of UE (score <2, *LS* group) were then invited to participate, following previous imaging research in HS (Premkumar *et al.*, 2012). The UE subscale of the O-LIFE was used as it is associated with higher severity of positive symptoms in schizophrenia (Cochrane *et al.*, 2010), and our first neuroanatomical study in schizotypy had also selected individuals based on positive-dimension scores on a similar self-report instrument (Modinos *et al.*, 2010).

Twenty-three individuals were included in the HS group (11 females; age range, 18–55 years; mean age 28.48 years) and 23 in the LS group (11 females; age range, 18–58 years; mean age 28.36 years). Participants with personal history of neurologic or psychiatric disorders were excluded, as assessed with the Mini International Neuropsychiatric Inventory (Sheehan *et al.*, 1998), administered by a trained interviewer. All participants were MRI-safe, and none had used recreational drugs in the two weeks prior to magnetic resonance imaging (MRI) scanning, or met criteria for substance abuse/dependency by self-report. Of note, there was no sample overlap between the current study and our previous volumetric study in HS (Modinos *et al.*, 2010). All participants gave written informed consent to the study protocol, which was approved by the King's College London Research Ethics Committee. The authors assert that all procedures contributing to this work comply with the ethical

standards of the relevant national and institutional committees on human experimentation and with the Helsinki Declaration of 1975, as revised in 2008.

Assessments

Subjects completed the following assessments before scanning commenced: a validated short version of the Wechsler Adult Intelligence Scale-III (WAIS-III) (Velthorst *et al.*, 2013); a semi-structured interview adapted from the Early Psychosis Prevention and Intervention Centre (EPPIC) Drug and Alcohol Assessment Schedule (<http://www.eppic.org.au>) to assess current/past medication use, current/past use of alcohol, tobacco, and illicit drugs; and the Social Functioning Questionnaire (SFQ) (Tyrer *et al.*, 2005).

Volumetric MRI Protocol

For all subjects, a three-dimensional T1-weighted inversion recovery prepared gradient echo sequence was obtained on a General Electric Discovery MR750 3T system (Milwaukee, WI, USA) at the Institute of Psychiatry, Psychology & Neuroscience, King's College London (voxel size: 1.05 x 1.05 x 1.2mm, field of view: 270mm, 196 slices, TR: 7.3 ms, TE: 3.0 ms, inversion time: 400 ms, flip angle = 11°, based on the well validated ADNI 2/ADNI GO protocols (see <http://adni.loni.usc.edu/methods/documents/mri-protocols/>).

Structural images were preprocessed using the Voxel-Based Morphometry protocol (Ashburner, 2010) implemented in SPM12 (<http://www.fil.ion.ucl.ac.uk/spm/software/spm12>), running on Matlab 9.2 (The MathWorks, USA). The following steps were applied: (1) Segmentation of all images into gray matter (GM), white matter (WM), and cerebrospinal fluid (CSF) partitions; (2) Warping of GM partitions into a new study-specific reference space, to provide increased accuracy of inter-subject alignment; (3) Normalization of the warped GM partitions to the MNI space, to

generate smoothed (10mm FWHM), spatially normalized and modulated GM images in Montreal Neuroanatomical Imaging (MNI) space. Global GM volume and total intracranial volume were calculated. Global GM volume was estimated using the total amount of gray matter in the GM images obtained from segmentation, including the cerebellar gray matter. Total intracranial volume was calculated for each subject by summing together the voxel values of GM, WM and CSF from the original tissue partitions using the ImCalc function in SPM12.

¹H-MRS Protocol and Quantification

A proton magnetic resonance spectrum (¹H-MRS; PRESS, Point RESolved Spectroscopy) was acquired during the same scanning session, after the structural MRI scan, from the bilateral anterior cingulate cortex (Figure 1A), as described in previous studies from our group in UHR subjects and patients with first-episode psychosis (Egerton *et al.*, 2012, Egerton *et al.*, 2014, Stone *et al.*, 2009), and schizotypy (Modinos *et al.*, 2017).

A standard GE PROBE (proton brain examination) sequence was used, which incorporates a standardized chemically selective suppression (CHESS) water suppression routine (TE: 30 ms; TR: 3000 ms; 96 averages collected). For each acquisition, unsuppressed water reference spectra (16 averages) were also acquired. Shimming was optimized, with auto-prescan performed twice before each scan. The region of interest (ROI) in the bilateral anterior cingulate cortex (ACC) was prescribed medially from the midline sagittal localizer, and the center of the 20 × 20 × 20 mm ROI mostly covering the ACC was placed 16 mm above the anterior section of the genu of the corpus callosum at 90° to the anterior commissure – posterior commissure (AC-PC) line.

Spectra were analyzed using LCModel version 6.3-1L (<http://s-provencher.com/pages/lcmodel.shtml>) (Provencher, 1993). Water-scaled glutamate (Glu), Glutamine (Gln), Glx (glutamate + glutamine), myo-Inositol (ml), choline (Cho), creatine (Cre) and N-acetylaspartate (NAA) values were corrected for voxel CSF using the formula: Metabolite Corrected = Metabolite Concentration * [proportion WM + (1.283 * proportion GM) + (1.548 * proportion CSF)] / (proportion WM + proportion GM) (Provencher, 2016). Voxel GM, WM and CSF content for each subject were ascertained by extracting the location of the voxel from the spectra file headers, and using an in-house program to calculate the percentage GM, WM and CSF using the segmented T1-weighted images. We used Cramer-Rao minimum variance bounds (CRLB) > 20% as reported by LCModel, which are estimates of fit of the metabolite peaks, and signal-to-noise ratio (SNR) <8 to exclude poorly-fitted metabolite peaks from statistical analysis (Mouchlianitis *et al.*, 2016, Provencher, 2016); only one HS subject exceeded the CRLB threshold and the final sample thus involved 22 HS subjects and 23 LS subjects. This sample largely overlaps with our recent publication in 21 HS and 22 LS subjects on functional MRI–MRS interactions (Modinos *et al.*, 2017). The primary ¹H-MRS measure was glutamate corrected for voxel CSF. No correction was made for T1- and T2-relaxation and metabolite concentrations are given in “institutional units”.

Statistics

Analysis of behavioral and demographic data was performed in SPSS 23 (<http://www-01.ibm.com/software/uk/analytics/spss/>). The effect of group on these measures was tested using independent samples *t*-tests for parametric data and Chi-square tests for non-parametric data. Significant effects are reported at $p < 0.05$.

¹H-MRS data analysis

Group differences in bilateral ACC glutamate concentrations were examined with an independent samples *t*-test in SPSS. Exploratory analyses of the other metabolites present in the spectra were also conducted with a *t*-test corrected for multiple comparisons (threshold for 6 metabolites, 1 voxel; $p = 0.008$). Levene's test was used to check for equality of variance across groups.

MRI data analysis

Statistical analyses of MRI data were implemented in SPM12 using an independent samples *t*-test, with proportional scaling of the total intracranial volume to adjust for global effects. We examined significant group effects in GM volume in HS and LS using a voxel-wise ROI approach. ROIs in the precuneus and posterior cingulate gyrus were specified *a-priori* using the coordinates from our previous structural study in an independent HS sample (Modinos *et al.*, 2010) and in the ACC from a previous UHR study (Borgwardt *et al.*, 2007a): MNI coordinates precuneus $x, y, z = 0, -54, 44$; posterior cingulate gyrus $x, y, z = 2, -42, 18$; and ACC $x, y, z = -6, 0, 43$. These coordinates were used for small volume correction (SVC) with a 10-mm sphere. For all tests, we used an initial voxel-wise search threshold of $p < 0.005$ uncorrected, to then only consider significant effects which survived voxel-wise family-wise error (FWE) correction for multiple comparisons at $p < 0.05$. Exploratory whole brain results were analyzed at the same initial uncorrected threshold for completeness, but only effects surviving voxel-wise $p < 0.05$ FWE correction are discussed.

Integration of MRI and ^1H -MRS data

The relationship between GM volume and glutamate levels in the ACC was established by using the individual glutamate concentrations as a regressor in a voxel-wise ROI ANOVA (precuneus, posterior and ACC as described above), also using proportional scaling of the

total intracranial volume to adjust for global effects. The same correction for multiple comparisons as described above was used.

Correlations between imaging and schizotypy / social functioning levels

Relationships between glutamate levels and both schizotypy scores and social functioning levels were explored in the HS group using Pearson's product moment correlation, correcting for the number of correlations performed (threshold $p < .05/2$ comparisons = .025). To assess correlations between GM volume and schizotypy scores / social functioning levels in HS individuals, two separate ROI regression analyses using the individual O-LIFE total / SFQ total scores as regressor in SPM12 were conducted, with the ROI coordinates as described above and applying the same correction for multiple comparisons.

RESULTS

Demographic and questionnaire data

Full demographic and behavioral results are presented in Table 1. The HS and LS groups only differed significantly by design in the schizotypy measures, with HS subjects showing higher scores than those with LS on the O-LIFE: Total ($p < .001$), Unusual Experiences ($p < .001$), Cognitive Disorganization ($p = .001$), and Introvertive Anhedonia ($p < .001$). High positive schizotypy (UE subscale) was positively associated with the CD ($r = .505$, $p = .017$) and marginally with the IA subscales ($r = .381$, $p = .080$).

[Table 1 about here]

¹H-MRS spectral quality

Spectra obtained were of good quality, with LCModel reporting mean \pm SD SNR of 25.6 ± 4.6 , line width of $4.9 \pm .8$ Hz and FWHM of $.04 \pm .004$. For Gln, only data from 9 LS subjects and 11 HS subjects were usable, with a similar number of subjects excluded from each group. There were no significant group differences in any of the parameters relating to spectral quality or voxel tissue content (Table 2).

Group differences in ^1H -MRS

There was no evidence of between-group differences in glutamate levels, or in any of the other metabolites present in the bilateral ACC voxel (glutamine, Glx, myo-inositol, choline, creatine, and N-acetylaspartate; Figure 1B; Table 2). There were no significant correlations between glutamate concentrations and age ($r = -.194$, $p = .203$) or caffeine use ($r = .189$, $p = .215$). Similarly, in the HS group no associations were found between GABA levels and social functioning scores ($r = .318$, $p = .149$), or O-LIFE total scores ($r = .107$, $p = .634$).

[Figure 1 about here]

[Table 2 about here]

Group differences in gray matter volume

Groups did not differ in global GM volume (LS, mean: $797.54 \pm 63.15 \text{ mm}^3$; HS, mean: $784.27 \pm 75.29 \text{ mm}^3$; $t = 0.642$; $p = 0.525$) or total intracranial volume (LS, mean: $1.0\text{e}+03*1.49 \pm .14 \text{ mm}^3$; HS, mean: $1.0\text{e}+03*1.50 \pm .11 \text{ mm}^3$; $t = -0.174$; $p = 0.862$).

Exploratory whole-brain analyses revealed smaller GM volume in HS compared to LS subjects in a right-sided region including the rolandic operculum and the superior temporal gyrus (peak coordinate at $x, y, z = 58 -20 14$; $T = 5.07$; $Z = 4.46$; $p_{FWE} = 0.045$) (Figure 2A).

There were no other significant whole-brain effects, nor significant effects were identified in our ROIs for the HS < LS contrast (voxel-wise FWE-corrected).

Using ROI analysis, subjects with HS showed larger GM volumes in the precuneus (peak coordinate at $x, y, z = -8 -50 46$; $T = 3.03$; $Z = 2.87$; $p_{FWE} = 0.043$) and in the ACC (peak coordinate at $x, y, z = -9 -8 39$; $T = 3.05$; $Z = 2.89$; $p_{FWE} = 0.041$) (Figure 2B). Groups did not differ in age or illicit drug/alcohol use, and the results of the GM volume comparisons did not change when age or illicit drug/alcohol use were added to the model. No regions survived voxel-wise FWE correction for the HS > LS contrast at the whole brain level.

ROI regression analyses were used to assess correlations between GM volume and schizotypal scores and social functioning. These revealed a positive association between O-LIFE total scores and GM volume in the precuneus (peak coordinate at $x, y, z = 0, -38, 24$; $T = 3.03$; $Z = 2.72$; $p_{FWE} = .044$), as well as positive association between SFQ total scores and precuneus volume (peak coordinate at $x, y, z = 2, -45, 12$; $T = 3.11$; $Z = 2.77$; $p_{FWE} = .039$). Thus, larger GM volumes in the precuneus were related to higher schizotypal traits and poorer social functioning in HS individuals.

[Figure 2 about here]

Relationship Between ^1H -MRS Measures and GMV

Figure 3 shows the relationship between glutamate levels and GM volume. In HS subjects, ROI analysis showed that ACC glutamate levels were negatively associated with local GM volume in the ACC. In the ACC, the higher the ACC glutamate, the smaller the volume of GM (peak coordinate at $x, y, z = -9, 9, 40$; $T = 3.28$; $Z = 3.07$; $p_{FWE} = 0.026$) (Figure 3). In terms of positive associations, no regions survived correction for multiple comparisons in HS. There

were no significant correlations in LS subjects in either directionality. Groups did not differ in age or illicit drug/alcohol use, and the results of the ACC glutamate-GM volume association did not change when age or illicit drug/alcohol use were added to the model. No effects survived voxel-wise FWE correction at the whole brain level.

A secondary analysis examining associations with levels of Glx showed a trend towards a negative association with GM volume in the ACC in HS individuals (peak coordinate at $x, y, z = -10, 8, 39$; $T = 2.96$; $Z = 2.79$ $p_{FWE} = .051$).

[Figure 3 about here]

DISCUSSION

The main aim of this study was to investigate whether glutamate levels are associated with gray matter volume in healthy individuals with high schizotypy, similar to the patterns observed in people at ultra-high risk of psychosis. Although glutamate concentrations did not significantly differ between individuals with high and low schizotypy, as we have recently reported in an overlapping dataset (Modinos *et al.*, 2017), glutamate levels in HS subjects were negatively associated with GM volume in the anterior cingulate cortex. This association was absent in the LS group. The findings are not attributable to effects of antipsychotics as all subjects were medication-naïve, and groups were matched for demographics, cognition, and substance use; the only significant differences between groups referred to the presence of subclinical psychotic-like experiences. This is consistent with evidence that levels of glutamate are negatively correlated with local GM volume in antipsychotic-naïve patients with first-episode psychosis (Kraguljac *et al.*, 2013, Plitman *et al.*, 2016). Our study extends those previous findings in caudate and hippocampus by showing the same relationship in

the ACC. Nevertheless, glutamate levels may also impact on more widespread structural changes, as reported in subjects at ultra-high risk of psychosis (Stone *et al.*, 2009) and patients with first-episode psychosis (Theberge *et al.*, 2007). Regardless of potentially disease-stage specific findings, our study suggests that ACC glutamate concentrations are related to volumetric changes in schizotypal individuals. Noteworthy, we recently reported that ACC glutamate concentrations were negatively correlated with neural response to emotion in brain regions robustly involved in the pathophysiology of psychosis (striatum and marginally MPFC and amygdala) in schizotypy (Modinos *et al.*, 2017). Our results suggest that ACC glutamate levels are related to increased local GM volume as well as to increased corticolimbic response to emotion in HS. Such relationships in HS provide some support to preclinical models postulating that altered prefronto-temporal-striatal interactions in relation to psychotic-like behaviors are associated with changes in cortical glutamatergic function (Berretta *et al.*, 2001, Lodge and Grace, 2011). As the normal function of this circuit depends on a proper excitation-inhibition balance, and given that the properties of this circuit appear to change during adolescence in preclinical models (Gomes *et al.*, 2016), our data provide some additional support for a “staged” approach to the understanding of the pathophysiology of schizophrenia-like symptoms and behaviors (Deutsch *et al.*, 2001). Collectively, these findings are in line with a psychosis continuum view by suggesting that interactions between brain structure and neurochemistry are involved in the expression of psychotic-like experiences at non-clinical and clinical degrees. These findings may also serve as evidence of potentially protective mechanisms, as this study involved high-functioning individuals with HS and considering that, while the association between ACC glutamate and GM volume was negative in HS, the group effect showed increased volume in this region compared to LS subjects. These results are opposite to the prediction from studies in clinical groups that HS would be associated with higher glutamate and reduced volume in that region. Further research examining the relative contributions of excitation and inhibition to

GM volume in different groups (schizophrenia, UHR, HS) and combining measurement of GABA and glutamate levels, may provide substantial insights into the neurobiology of risk and resilience for psychiatric disorders (Pantelis and Bartholomeusz, 2014).

The present study replicated our previous finding of larger regional GM volumes in the posterior cortical midline (Modinos *et al.*, 2010) in a separate cohort of HS individuals. We again report that people with HS had increased volume of the precuneus relative to comparison subjects with low levels of schizotypy, an area involved in self-awareness and self-evaluation and which is a core component of the default-mode network (Cavanna and Trimble, 2006, Utevsky *et al.*, 2014). Furthermore, larger precuneus volumes were significantly related to both higher levels of schizotypy as well as to lower social functioning in the HS sample, providing a link between these neuroanatomical findings and psychosis-related behaviors. A similar finding was reported by another study showing a positive association between precuneus volume and schizotypy scores (Nenadic *et al.*, 2015). In contrast, GM decreases as opposed to increases in this region were found by other groups in people at UHR of psychosis (Borgwardt *et al.*, 2007b) and patients with a first episode of psychosis (Theberge *et al.*, 2007). Although these findings, taken together, may suggest that volumetric changes in the precuneus are present along the psychosis continuum, the discrepancies in directionality indicate that structural characteristics of this region may vary based on the degree of psychosis risk (larger volumes in healthy people with schizotypy, at low risk) and expression (smaller volumes in people at higher risk or clinically ill). The functional significance of larger precuneus volumes associated with schizotypal traits, in particular, remains to be determined. One possibility is that such larger volumes, while associated with some of the subclinical features of psychosis, may offer resilience to the development of a clinical diagnosis. While the mechanisms are unclear, one speculative interpretation is that this may relate to variables such as creativity and spirituality, on which

people with schizotypal traits score higher on average. Indeed, creativity has been associated with larger precuneus volume and thickness (Chen *et al.*, 2015) and spirituality had also been associated with larger precuneus volume (Miller *et al.*, 2014). Higher levels of spirituality and greater precuneus volumes may confer resilience to developing depression (Miller *et al.*, 2014), and it is possible that larger volumes may also confer resilience to developing psychosis. A further explanation may be that these neuroanatomical changes reflect plastic adaptation to a genetic- or environment-related liability for psychiatric outcomes, and enable psychiatrically resilient high schizotypes to effectively compensate, in line with recent postulations about the effects of early life-stress on brain (Teicher *et al.*, 2016). The possibility that such larger volumes may offer resilience to the development of clinical psychosis, and the relationship with stress, deserves further investigation.

Further group effects in our study involved GM volume increases in anterior cingulate gyrus and decreases in superior temporal cortex. Volumetric decreases in superior temporal cortex areas have been extensively associated with schizophrenia (Honea *et al.*, 2005, Radua *et al.*, 2012), individuals at UHR of psychosis (Borgwardt *et al.*, 2008, Fusar-Poli *et al.*, 2011), and HS (Ettinger *et al.*, 2012). In the anterior cingulate gyrus, however, our study highlights an additional discrepancy as previous studies had mainly reported volumetric decreases. Increases in GM volume have been suggested to occur in the initial stages of apoptosis (Adler *et al.*, 2005), a process in which glutamatergic dysfunction is thought to be involved, so one possibility is that volumetric changes across the psychosis spectrum are dynamic, and that volumetric reductions occur at a later stage when a glutamatergic alteration is identifiable. However, no studies have examined the relationship between such structural findings and metabolite measures in relation to subclinical psychotic-like experiences, and our study suggests a direct link between them in the anterior cingulate cortex.

There are some limitations to the present study. Firstly, subjects were recruited from a university sample, had relatively high IQs and groups showed no differences in self-reported substance use; therefore, our results may not generalize to all individuals with schizotypy. Larger studies in general population samples would help define the normal variation in schizotypy. Secondly, several theories exist about the schizotypy construct and a range of assessment instruments have been developed; the present investigation used the O-LIFE questionnaire, which follows a personality-based model of schizotypy, and therefore should be interpreted to report on neurobiological correlates of schizotypal traits as understood from a dimensional rather than a clinical perspective. Thirdly, schizotypy is regarded as a multidimensional construct (Mason and Claridge, 2006, Nelson *et al.*, 2013). While the HS group was formed based on high UE scores, thus reflecting high positive schizotypy, groups also differed in the CD and IA subscales; this might have contributed to the present results and future studies should seek to explicitly explore the other schizotypy dimensions. Furthermore, this cross-sectional study does not allow evaluation of whether the observed GM changes are driven by HS subjects who may later develop a mental health disorder (Cannon *et al.*, 2015). With regards to MRS methodology, ^1H -MRS measurements of glutamate and glutamine reflect the combined intra- and extra-neuronal metabolite concentration, and using our approach of PRESS with a TE of 30 ms at 3 Tesla the partially overlapping signals from glutamate and glutamine cannot be entirely resolved, with contamination of the Glu signal by Gln estimated as <10% (Snyder and Wilman, 2010). Of note, groups did not differ significantly in either glutamine or Glx. This aligns with recent results showing that chronicity of psychotic symptoms is associated with decreased levels of Glx in a study comparing chronic schizophrenia patients, patients with recent onset psychotic disorder, and ultra-high risk individuals (Liemburg *et al.*, 2016) and suggests that detectable ^1H -MRS differences in non-clinical groups may be very subtle. We cannot rule out

the possibility that the lack of significant differences in the standalone ^1H -MRS measures are related to limited power in this non-clinical sample and this will need to be replicated by independent studies of larger samples. Finally, we employed a self-report of use of drugs of abuse but future studies should consider adding a drug screening test prior to scanning.

The present study suggests that gray matter volume changes in cortical midline areas are associated with glutamate levels in subjects with high schizotypy, with such findings not being dependent on disease chronicity or the confounding influence of antipsychotic medication. Given that our sample consisted of high-functioning schizotypes, these results may also indicate potential neurobiological mechanisms of resilience. Future longitudinal studies testing this model comprehensively, including serial structural imaging scans in the same individuals in combination with GABAergic, glutamatergic and dopaminergic neurotransmission, will be fundamental to understanding the mechanisms underlying the development of schizophrenia, and may provide a scientific basis for the development of novel interventions including facilitation of NMDA receptor-mediated neurotransmission to prevent or delay progression to psychosis in vulnerable individuals.

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Conflict of interest

GJB received honoraria for teaching from General Electric Healthcare, and acted as a consultant for IXICO, at the time of this study. The other authors declare no competing financial interests.

REFERENCES

- Abbott, C. & Bustillo, J.** (2006). What have we learned from proton magnetic resonance spectroscopy about schizophrenia? A critical update. *Current Opinion in Psychiatry* **19**, 135-9.
- Adler, C. M., Levine, A. D., DelBello, M. P. & Strakowski, S. M.** (2005). Changes in gray matter volume in patients with bipolar disorder. *Biological Psychiatry* **58**, 151-7.
- Aoyama, N., Theberge, J., Drost, D. J., Manchanda, R., Northcott, S., Neufeld, R. W., Menon, R. S., Rajakumar, N., Pavlosky, W. F., Densmore, M., Schaefer, B. & Williamson, P. C.** (2011). Grey matter and social functioning correlates of glutamatergic metabolite loss in schizophrenia. *The British Journal of Psychiatry* **198**, 448-56.
- Ashburner, J.** (2010). VBM Tutorial. Available at: <http://www.fil.ion.ucl.ac.uk/~john/misc/VBMclass10.pdf>.
- Barrantes-Vidal, N., Grant, P. & Kwapil, T. R.** (2015). The role of schizotypy in the study of the etiology of schizophrenia spectrum disorders. *Schizophrenia Bulletin* **41 Suppl 2**, S408-16.
- Berretta, S., Munno, D. W. & Benes, F. M.** (2001). Amygdalar activation alters the hippocampal GABA system: "partial" modelling for postmortem changes in schizophrenia. *Journal of Comparative Neurology* **431**, 129-38.
- Boos, H. B., Aleman, A., Cahn, W., Hulshoff Pol, H. & Kahn, R. S.** (2007). Brain volumes in relatives of patients with schizophrenia: a meta-analysis. *Archives of General Psychiatry* **64**, 297-304.
- Bora, E., Fornito, A., Radua, J., Walterfang, M., Seal, M., Wood, S. J., Yucel, M., Velakoulis, D. & Pantelis, C.** (2011). Neuroanatomical abnormalities in schizophrenia: a multimodal voxelwise meta-analysis and meta-regression analysis. *Schizophrenia Research* **127**, 46-57.
- Borgwardt, S. J., McGuire, P., Fusar-Poli, P., Radue, E. W. & Riecher-Rossler, A.** (2008). Anterior cingulate pathology in the prodromal stage of schizophrenia. *Neuroimage* **39**, 553-4.
- Borgwardt, S. J., McGuire, P. K., Aston, J., Berger, G., Dazzan, P., Gschwandtner, U., Pfluger, M., D'Souza, M., Radue, E. W. & Riecher-Rossler, A.** (2007a). Structural brain abnormalities in individuals with an at-risk mental state who later develop psychosis. *The British Journal of Psychiatry Supplement* **51**, s69-75.
- Borgwardt, S. J., Riecher-Rossler, A., Dazzan, P., Chitnis, X., Aston, J., Drewe, M., Gschwandtner, U., Haller, S., Pfluger, M., Rechsteiner, E., D'Souza, M., Stieglitz, R. D., Radu, E. W. & McGuire, P. K.** (2007b). Regional gray matter volume abnormalities in the at risk mental state. *Biological Psychiatry* **61**, 1148-56.
- Cannon, T. D., Chung, Y., He, G., Sun, D., Jacobson, A., van Erp, T. G., McEwen, S., Addington, J., Bearden, C. E., Cadenhead, K., Cornblatt, B., Mathalon, D. H., McGlashan, T., Perkins, D., Jeffries, C., Seidman, L. J., Tsuang, M., Walker, E., Woods, S. W., Heinssen, R. & North American Prodrome Longitudinal Study, C.** (2015). Progressive reduction in cortical thickness as psychosis develops: a multisite longitudinal neuroimaging study of youth at elevated clinical risk. *Biological Psychiatry* **77**, 147-57.
- Cavanna, A. E. & Trimble, M. R.** (2006). The precuneus: a review of its functional anatomy and behavioural correlates. *Brain* **129**, 564-83.
- Chan, R. C., Di, X., McAlonan, G. M. & Gong, Q. Y.** (2011). Brain anatomical abnormalities in high-risk individuals, first-episode, and chronic schizophrenia: an activation likelihood estimation meta-analysis of illness progression. *Schizophrenia Bulletin* **37**, 177-88.
- Chen, Q. L., Xu, T., Yang, W. J., Li, Y. D., Sun, J. Z., Wang, K. C., Beaty, R. E., Zhang, Q. L., Zuo, X. N. & Qiu, J.** (2015). Individual differences in verbal creative thinking are reflected in the precuneus. *Neuropsychologia* **75**, 441-9.
- Choi, D. W.** (1994). Calcium and excitotoxic neuronal injury. *Annals of the New York Academy of Sciences* **747**, 162-71.

- Cochrane, M., Petch, I. & Pickering, A. D.** (2010). Do measures of schizotypal personality provide non-clinical analogues of schizophrenic symptomatology? *Psychiatry Research* **176**, 150-4.
- Deutsch, S. I., Rosse, R. B., Schwartz, B. L. & Mastropalo, J.** (2001). A revised excitotoxic hypothesis of schizophrenia: therapeutic implications. *Clinical Neuropharmacology* **24**, 43-9.
- Egerton, A., Bhachu, A., Merritt, K., McQueen, G., Szulc, A. & McGuire, P.** (2017). Effects of Antipsychotic Administration on Brain Glutamate in Schizophrenia: A Systematic Review of Longitudinal 1H-MRS Studies. *Frontiers in Psychiatry* **8**.
- Egerton, A., Brugger, S., Raffin, M., Barker, G. J., Lythgoe, D. J., McGuire, P. K. & Stone, J. M.** (2012). Anterior cingulate glutamate levels related to clinical status following treatment in first-episode schizophrenia. *Neuropsychopharmacology* **37**, 2515-21.
- Egerton, A., Stone, J. M., Chaddock, C. A., Barker, G. J., Bonoldi, I., Howard, R. M., Merritt, K., Allen, P., Howes, O. D., Murray, R. M., McLean, M. A., Lythgoe, D. J., O'Gorman, R. L. & McGuire, P. K.** (2014). Relationship Between Brain Glutamate Levels and Clinical Outcome in Individuals at Ultra High Risk of Psychosis. *Neuropsychopharmacology* **39**, 2891-9.
- Ettinger, U., Williams, S. C., Meisenzahl, E. M., Moller, H. J., Kumari, V. & Koutsouleris, N.** (2012). Association between brain structure and psychometric schizotypy in healthy individuals. *The World Journal of Biological Psychiatry* **13**, 544-9.
- Fusar-Poli, P., Borgwardt, S., Crescini, A., Deste, G., Kempton, M. J., Lawrie, S., McGuire, P. & Sacchetti, E.** (2011). Neuroanatomy of vulnerability to psychosis: a voxel-based meta-analysis. *Neuroscience & Biobehavioral Reviews* **35**, 1175-85.
- Gomes, F. V., Rincon-Cortes, M. & Grace, A. A.** (2016). Adolescence as a period of vulnerability and intervention in schizophrenia: Insights from the MAM model. *Neuroscience & Biobehavioral Reviews* **70**, 260-70.
- Haijma, S. V., Van Haren, N., Cahn, W., Koolschijn, P. C., Hulshoff Pol, H. E. & Kahn, R. S.** (2013). Brain volumes in schizophrenia: a meta-analysis in over 18 000 subjects. *Schizophrenia Bulletin* **39**, 1129-38.
- Honea, R., Crow, T. J., Passingham, D. & Mackay, C. E.** (2005). Regional deficits in brain volume in schizophrenia: a meta-analysis of voxel-based morphometry studies. *The American Journal of Psychiatry* **162**, 2233-45.
- Kraguljac, N. V., White, D. M., Reid, M. A. & Lahti, A. C.** (2013). Increased hippocampal glutamate and volumetric deficits in unmedicated patients with schizophrenia. *JAMA Psychiatry* **70**, 1294-302.
- Kwapil, T. R., Gross, G. M., Silvia, P. J. & Barrantes-Vidal, N.** (2013). Prediction of psychopathology and functional impairment by positive and negative schizotypy in the Chapmans' ten-year longitudinal study. *Journal of Abnormal Psychology* **122**, 807-15.
- Liemburg, E., Sibeijn-Kuiper, A., Bais, L., Pijnenborg, G., Knegtering, H., van der Velde, J., Opmeer, E., de Vos, A., Dlabac-De Lange, J., Wunderink, L. & Aleman, A.** (2016). Prefrontal NAA and Glx Levels in Different Stages of Psychotic Disorders: a 3T 1H-MRS Study. *Scientific Reports* **6**, 21873.
- Linscott, R. J. & van Os, J.** (2013). An updated and conservative systematic review and meta-analysis of epidemiological evidence on psychotic experiences in children and adults: on the pathway from proneness to persistence to dimensional expression across mental disorders. *Psychological Medicine* **43**, 1133-49.
- Lodge, D. J. & Grace, A. A.** (2011). Hippocampal dysregulation of dopamine system function and the pathophysiology of schizophrenia. *Trends in Pharmacological Sciences* **32**, 507-13.
- Marsman, A., van den Heuvel, M. P., Klomp, D. W., Kahn, R. S., Luijten, P. R. & Hulshoff Pol, H. E.** (2013). Glutamate in schizophrenia: a focused review and meta-analysis of (1)H-MRS studies. *Schizophrenia Bulletin* **39**, 120-9.
- Mason, O. & Claridge, G.** (2006). The Oxford-Liverpool Inventory of Feelings and Experiences (O-LIFE): further description and extended norms. *Schizophrenia Research* **82**, 203-11.

- Mason, O., Linney, Y. & Claridge, G.** (2005). Short scales for measuring schizotypy. *Schizophrenia Research* **78**, 293-6.
- McDonald, C., Dineen, B. & Hallahan, B.** (2008). Meta-analysis of brain volumes in unaffected first-degree relatives of patients with schizophrenia overemphasizes hippocampal deficits. *Archives of General Psychiatry* **65**, 603-4; author reply 604-5.
- Mechelli, A., Riecher-Rossler, A., Meisenzahl, E. M., Tognin, S., Wood, S. J., Borgwardt, S. J., Koutsouleris, N., Yung, A. R., Stone, J. M., Phillips, L. J., McGorry, P. D., Valli, I., Velakoulis, D., Woolley, J., Pantelis, C. & McGuire, P.** (2011). Neuroanatomical abnormalities that predate the onset of psychosis: a multicenter study. *Archives of General Psychiatry* **68**, 489-95.
- Merritt, K., Egerton, A., Kempton, M. J., Taylor, M. J. & McGuire, P. K.** (2016). Nature of Glutamate Alterations in Schizophrenia: A Meta-analysis of Proton Magnetic Resonance Spectroscopy Studies. *JAMA Psychiatry* **73**, 665-674.
- Miller, L., Bansal, R., Wickramaratne, P., Hao, X., Tenke, C. E., Weissman, M. M. & Peterson, B. S.** (2014). Neuroanatomical correlates of religiosity and spirituality: a study in adults at high and low familial risk for depression. *JAMA Psychiatry* **71**, 128-35.
- Modenato, C. & Draganski, B.** (2015). The concept of schizotypy — A computational anatomy perspective. *Schizophrenia Research: Cognition* **2**, 89-92.
- Modinos, G., McLaughlin, A., Egerton, A., McMullen, K., Kumari, V., Barker, G. J., Keyzers, C. & Williams, S. C.** (2017). Corticolimbic hyper-response to emotion and glutamatergic function in people with high schizotypy: a multimodal fMRI-MRS study. *Translational Psychiatry* **7**, e1083.
- Modinos, G., Mechelli, A., Ormel, J., Groenewold, N. A., Aleman, A. & McGuire, P. K.** (2010). Schizotypy and brain structure: a voxel-based morphometry study. *Psychological Medicine* **40**, 1423-31.
- Mouchlianitis, E., Bloomfield, M. A., Law, V., Beck, K., Selvaraj, S., Rasquinha, N., Waldman, A., Turkheimer, F. E., Egerton, A., Stone, J. & Howes, O. D.** (2016). Treatment-Resistant Schizophrenia Patients Show Elevated Anterior Cingulate Cortex Glutamate Compared to Treatment-Responsive. *Schizophrenia Bulletin* **42**, 744-52.
- Nelson, M. T., Seal, M. L., Pantelis, C. & Phillips, L. J.** (2013). Evidence of a dimensional relationship between schizotypy and schizophrenia: a systematic review. *Neuroscience & Biobehavioral Reviews* **37**, 317-27.
- Nenadic, I., Lorenz, C., Langbein, K., Dietzek, M., Smesny, S., Schonfeld, N., Fananas, L., Sauer, H. & Gaser, C.** (2015). Brain structural correlates of schizotypy and psychosis proneness in a non-clinical healthy volunteer sample. *Schizophrenia Research* **168**, 37-43.
- Pantelis, C. & Bartholomeusz, C. F.** (2014). Social neuroscience in psychiatry: pathways to discovering neurobiological risk and resilience. *World Psychiatry* **13**, 146-7.
- Plitman, E., Patel, R., Chung, J. K., Pipitone, J., Chavez, S., Reyes-Madriral, F., Gomez-Cruz, G., Leon-Ortiz, P., Chakravarty, M. M., de la Fuente-Sandoval, C. & Graff-Guerrero, A.** (2016). Glutamatergic Metabolites, Volume and Cortical Thickness in Antipsychotic-Naive Patients with First-Episode Psychosis: Implications for Excitotoxicity. *Neuropsychopharmacology* **41**, 2606-13.
- Port, J. D. & Agarwal, N.** (2011). MR spectroscopy in schizophrenia. *Journal of Magnetic Resonance Imaging* **34**, 1251-61.
- Premkumar, P., Ettinger, U., Inchley-Mort, S., Sumich, A., Williams, S. C., Kuipers, E. & Kumari, V.** (2012). Neural processing of social rejection: the role of schizotypal personality traits. *Human Brain Mapping* **33**, 695-706.
- Provencher, S. W.** (1993). Estimation of metabolite concentrations from localized in vivo proton NMR spectra. *Magnetic Resonance in Medicine* **30**, 672-9.
- Provencher, S. W.** (2016). LCMoDel & LCMgui User's Manual.

- Radua, J., Borgwardt, S., Crescini, A., Mataix-Cols, D., Meyer-Lindenberg, A., McGuire, P. K. & Fusar-Poli, P.** (2012). Multimodal meta-analysis of structural and functional brain changes in first episode psychosis and the effects of antipsychotic medication. *Neuroscience & Biobehavioral Reviews* **36**, 2325-33.
- Raine, A.** (1991). The SPQ: a scale for the assessment of schizotypal personality based on DSM-III-R criteria. *Schizophrenia Bulletin* **17**, 555-64.
- Satterthwaite, T. D., Wolf, D. H., Calkins, M. E., Vandekar, S. N., Erus, G., Ruparel, K., Roalf, D. R., Linn, K. A., Elliott, M. A., Moore, T. M., Hakonarson, H., Shinohara, R. T., Davatzikos, C., Gur, R. C. & Gur, R. E.** (2016). Structural Brain Abnormalities in Youth With Psychosis Spectrum Symptoms. *JAMA Psychiatry* **73**, 515-24.
- Sheehan, D. V., Lecrubier, Y., Sheehan, K. H., Amorim, P., Janavs, J., Weiller, E., Hergueta, T., Baker, R. & Dunbar, G. C.** (1998). The Mini-International Neuropsychiatric Interview (M.I.N.I.): the development and validation of a structured diagnostic psychiatric interview for DSM-IV and ICD-10. *Journal of Clinical Psychiatry* **59 Supplement 20**, 22-33;quiz 34-57.
- Snyder, J. & Wilman, A.** (2010). Field strength dependence of PRESS timings for simultaneous detection of glutamate and glutamine from 1.5 to 7T. *Journal of Magnetic Resonance* **203**, 66-72.
- Stone, J. M., Day, F., Tsagaraki, H., Valli, I., McLean, M. A., Lythgoe, D. J., O'Gorman, R. L., Barker, G. J., McGuire, P. K. & Oasis** (2009). Glutamate dysfunction in people with prodromal symptoms of psychosis: relationship to gray matter volume. *Biological Psychiatry* **66**, 533-9.
- Teicher, M. H., Samson, J. A., Anderson, C. M. & Ohashi, K.** (2016). The effects of childhood maltreatment on brain structure, function and connectivity. *Nature Reviews Neuroscience* **17**, 652-66.
- Theberge, J., Williamson, K. E., Aoyama, N., Drost, D. J., Manchanda, R., Malla, A. K., Northcott, S., Menon, R. S., Neufeld, R. W., Rajakumar, N., Pavlosky, W., Densmore, M., Schaefer, B. & Williamson, P. C.** (2007). Longitudinal grey-matter and glutamatergic losses in first-episode schizophrenia. *The British Journal of Psychiatry* **191**, 325-34.
- Tyrer, P., Nur, U., Crawford, M., Karlsen, S., McLean, C., Rao, B. & Johnson, T.** (2005). The Social Functioning Questionnaire: a rapid and robust measure of perceived functioning. *International Journal of Social Psychiatry* **51**, 265-75.
- Utevsky, A. V., Smith, D. V. & Huettel, S. A.** (2014). Precuneus is a functional core of the default-mode network. *The Journal of Neuroscience* **34**, 932-40.
- Velthorst, E., Levine, S. Z., Henquet, C., de Haan, L., van Os, J., Myin-Germeys, I. & Reichenberg, A.** (2013). To cut a short test even shorter: reliability and validity of a brief assessment of intellectual ability in schizophrenia--a control-case family study. *Cognitive Neuropsychiatry* **18**, 574-93.
- Wang, Y., Yan, C., Yin, D. Z., Fan, M. X., Cheung, E. F., Pantelis, C. & Chan, R. C.** (2015). Neurobiological changes of schizotypy: evidence from both volume-based morphometric analysis and resting-state functional connectivity. *Schizophrenia Bulletin* **41 Suppl 2**, S444-54.
- Wiebels, K., Waldie, K. E., Roberts, R. P. & Park, H. R.** (2016). Identifying grey matter changes in schizotypy using partial least squares correlation. *Cortex* **81**, 137-50.

FIGURE LEGENDS

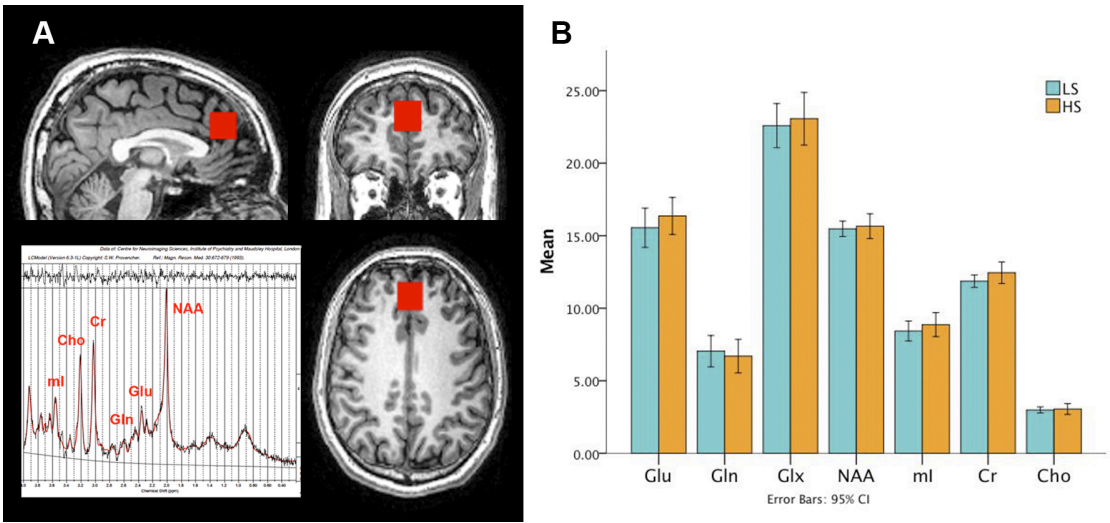


Figure 1. (A) Magnetic Resonance Spectroscopy voxel placement in the anterior cingulate cortex. (B) Bar graphs showing metabolite levels (in institutional units) in each group (LS, low schizotypy; HS = high schizotypy). Glu = glutamate; Gln = glutamine; Glx = glutamate + glutamine; NAA = N-acetylaspartate; Mi = myo-inositol; Cr = creatine; Cho = choline.

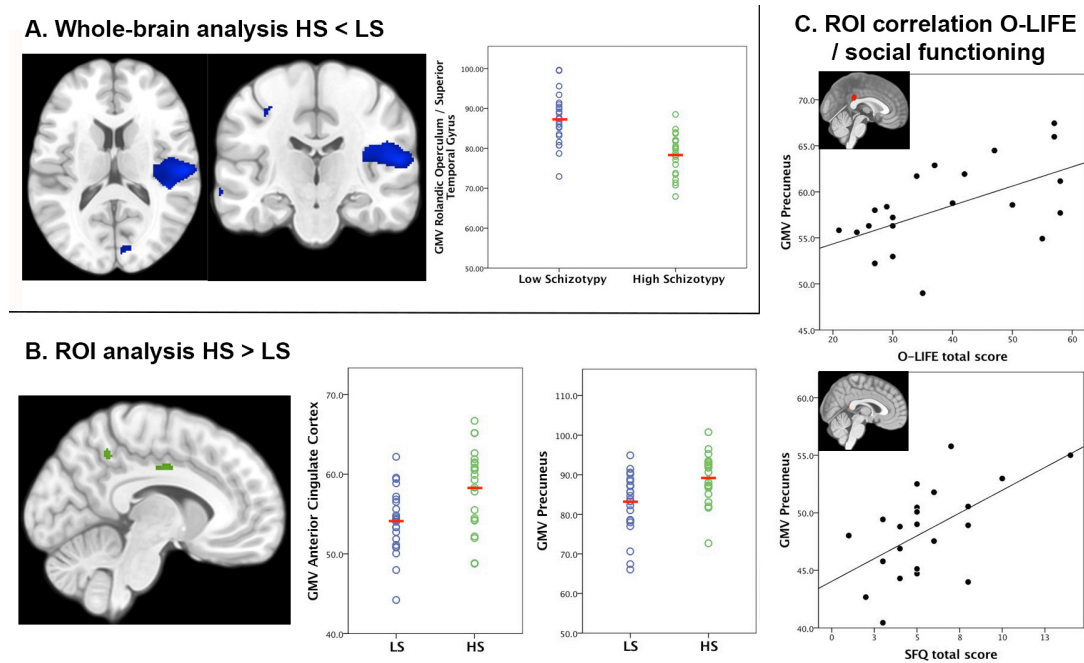


Figure 2. (A) Decreased gray matter volume (GMV) in the rolandic operculum/superior temporal gyrus in subjects high schizotypy (HS) compared to those with low schizotypy (LS). (B) Increased GMV in the precuneus and the anterior cingulate cortex in HS compared to LS subjects. (C) Positive associations between GMV in the precuneus and levels of schizotypy (O-LIFE total, upper panel) and levels of social functioning (SFQ total, lower panel). Higher O-LIFE scores indicate higher schizotypy, and higher SFQ scores indicate poorer social functioning. All effects were considered significant at voxel-wise $P < .05$ FWE correction; statistical parametric maps thresholded at $P < .005$ uncorrected for display purposes.

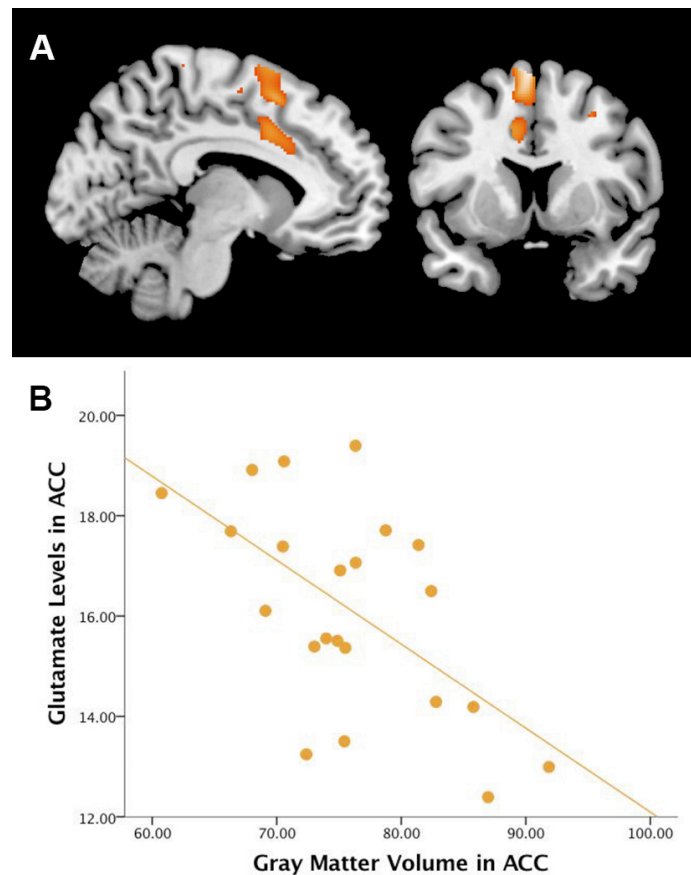


Figure 3. (A) Section overlay of the negative correlation between glutamate levels and gray matter volume in the anterior cingulate cortex (ACC) in high schizotypy subjects. Effects were considered significant at voxel-wise $P < .05$ FWE correction; statistical parametric maps are shown at $P < .005$ uncorrected for display purposes. (B) Plot depicting the negative correlation between glutamate levels and grey matter volume in the ACC.

TABLES

Table 1. Sample Characteristics.

	Low Schizotypy (<i>n</i> = 23)	High Schizotypy (<i>n</i> = 22)	Analysis
Age (years)	26.87 (5.40)	27.36 (7.61)	$t = -.252$ $P = .802$
Gender (% female)	47.8% (<i>n</i> = 11)	50% (<i>n</i> = 11)	$\chi^2 = .021$ $P = .884$
Ethnicity (% white)	73.9% (<i>n</i> = 17)	59.1% (<i>n</i> = 13)	$\chi^2 = 4.604$ $P = .203$
IQ (WAIS-III short version)	122.30 (13.54)	117.41 (18.55)	$t = 1.014$ $P = .316$
O-LIFE Total	16.04 (8.58)	38.23 (12.62)	$t = -6.865$ $P < .001$
O-LIFE Unusual Experiences	.96 (1.02)	11.59 (4.93)	$t = -9.926$ $P < .001$
O-LIFE Cognitive Disorganization	5.48 (3.96)	11.32 (6.69)	$t = -3.544$ $P = .001$
O-LIFE Introvertive Anhedonia	4.78 (3.18)	9.05 (2.50)	$t = -4.990$ $P < .001$
O-LIFE Impulsive Nonconformity	4.83 (4.24)	6.27 (4.60)	$t = -1.099$ $P = .278$
Social Functioning Questionnaire Total	4.09 (3.04)	5.50 (2.87)	$t = -1.600$ $P = .117$
Daily tobacco use (mean)	.71 (3.19)	.30 (0.75)	$t = .552$ $P = .584$
Daily caffeine use (mean)	1.78 (1.46)	2.82 (2.52)	$t = -1.752$ $P = .087$
Alcohol use (median [range])	2 (0 – 5)	1 (0 – 4)	$\chi^2 = 6.047$ $P = .302$
Marijuana use (median [range])	.5 (0 – 3)	0 (0 – 3)	$\chi^2 = 1.767$ $P = .622$
Parental socio-economic status (% professional level)	63.6% (<i>n</i> = 14)	70.0% (<i>n</i> = 14)	$\chi^2 = .573$ $P = .751$
Educational level (% university level)	91.3% (<i>n</i> = 21)	77.3% (<i>n</i> = 17)	$\chi^2 = 1.685$ $P = .194$

Note: higher scores on the Social Functioning Questionnaire reflect lower social functioning.
O-LIFE = Oxford–Liverpool Inventory of Feelings and Experiences; Wechsler Adult Intelligence Scale-III.

Table 2. ¹H-MRS quality parameters and metabolite levels by group.

	Low Schizotypy		High Schizotypy		P-value
	Mean	SD	Mean	SD	
SNR	25.26	4.34	25.73	4.76	.733
Line width	5.04	1.04	4.72	0.56	.199
FWHM	0.038	0.004	0.036	0.005	.275
Voxel CSF	0.24	0.04	0.26	0.05	.405
Voxel GM	0.64	0.05	0.63	0.05	.429
Voxel WM	0.11	0.03	0.11	0.04	.937
Glutamate % CRLB	6.17	1.40	5.73	0.94	.218
Glutamate	15.67	2.51	16.14	2.07	.497
Glutamine % CRLB	14.44 (n=9)	4.07	14.73 (n=11)	3.50	.869
Glutamine	7.04 (n=9)	1.41	6.70 (n=11)	1.72	.642
Glx % CRLB	6.87	2.10	6.32	1.21	.289
Glx	20.36	3.67	21.27	3.48	.398
Creatine % CRLB	2.43	0.51	2.50	0.51	.670
Creatine	12.15	1.45	12.05	1.17	.805
Myo-Inositol % CRLB	5.17	2.23	4.64	0.90	.299
Myo-Inositol	8.54	1.60	8.72	1.05	.652
NAA % CRLB	2.96	0.56	2.73	0.46	.141
NAA	14.88	1.34	15.25	1.54	.396
Choline % CRLB	3.13	0.34	3.09	0.53	.766
Choline	2.94	0.36	2.99	0.46	.699

SNR: signal-to-noise ratio; CRLB: Cramer-Rao Lower Bounds, CSF: cerebrospinal fluid; FWHM: full-width-half-maximum; GM: gray matter; NAA: N-acetylaspartate; WM: white matter.